

Original Article

Prognostic value of leukocyte telomere length in renal cell carcinoma patients

Meng Chen^{1,2}, Chia-Wen Tsai^{2,3}, Wen-Shin Chang^{2,3}, Junfeng Xu², Yifan Xu², Da-Tian Bau^{3,4*}, Jian Gu^{2*}

¹Department of Clinical Laboratory, National Cancer Center/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China; ²Department of Epidemiology, The University of Texas MD Anderson Cancer Center, Houston, TX 77030, USA; ³Terry Fox Cancer Research Laboratory, China Medical University Hospital, Taichung, Taiwan; ⁴Department of Bioinformatics and Medical Engineering, Asia University, Taichung, Taiwan. *Equal contributors.

Received June 30, 2020; Accepted July 9, 2020; Epub October 1, 2020; Published October 15, 2020

Abstract: Telomeres play important roles in cancer initiation and progression. Leukocyte telomere length (LTL) can modulate cancer risk and outcome. We hypothesize that genetically predicted short LTL is associated with worse prognosis in renal cell carcinoma (RCC). A total of 1,086 histologically confirmed RCC patients were included in this study. A weighted genetic risk score (GRS) predictive of LTL was constructed using 10 confirmed LTL-associated single nucleotide polymorphisms (SNPs). The associations of individual SNPs and GRS with recurrence and survival were determined by multivariate Cox proportional hazards analysis. In individual SNP analysis, long LTL-associated allele of rs7675998 in NAF1 gene at chromosome 4 was significantly associated with a reduced risk of recurrence (HR=0.85, 95% CI, 0.73-0.99, P=0.043), while the long LTL-associated allele of rs10936599 in TERC on chromosome 3 conferred a reduced risk of death (HR=0.85, 95% CI, 0.73-1.00, P=0.047). More importantly, genetically predicted LTL was associated with both recurrence and survival. Dichotomized at the median value of GRS, patients with low GRS (indicating short LTL) exhibited significantly increased risks of recurrence (HR=1.26, 95% CI, 1.03-1.54, P=0.025) and death (HR=1.23, 95% CI, 1.00-1.50, P=0.045). Hence, we concluded that genetically predicted short LTL is associated with worse prognosis in RCC patients.

Keywords: Renal cell carcinoma, leukocyte telomere length, recurrence, survival, genetic risk score (GRS), Mendelian randomization

Introduction

Renal cell carcinoma (RCC) accounts for about 85% of adult kidney cancers and is the most lethal genitourinary cancer. The incidence of RCC has been steadily rising by 2-4% per year in the past four decades until recent years when the incidence rate stabilized [1, 2]. About two thirds of patients with RCC have localized diseases at diagnosis and the remaining one third of patients present with regional and distant metastatic RCC [1]. The clinical management of RCC has evolved significantly in the past decade when novel surgical and systemic therapies have improved the prognosis of RCC [3]. Although patients with localized RCC can be cured by nephrectomy, about 40% of patients will develop recurrence/metastasis after surgical resection and eventually die from

this disease [3]. The success of targeted and immune checkpoint therapies in metastatic RCC has generated intense interest in using these novel therapies in adjuvant setting to prevent diseases recurrence/metastasis. To improve prognosis of surgically resected RCC, effective adjuvant therapy is clearly needed for high risk RCC. However, to date, the only FDA approved systemic adjuvant therapy for RCC is sunitinib, which offered a 2-month disease-free survival benefit accompanied by significant drug-related toxicities [4]. Many clinical trials of targeted and immune therapy for localized, high-risk RCC have been completed or are ongoing [5, 6]. Whereas efficacy has been mostly disappointing, toxicity has been consistently a major concern. Patient selection based on accurate risk stratification algorithm is obviously a pre-requisite for successful adjuvant

therapy. There are several clinically used nomograms for predicting recurrence risk in surgically resected RCC patients that rely solely on clinicopathologic variables (e.g., histology, tumor size, TNM stage, Fuhrman grade, and performance status) [5], but these clinical variable-based prognostic models are not sufficient. Identifying independent biomarkers that can supplement clinical variables to determine which patients are most likely to recur/metastasize after surgery has tremendous clinical value [7, 8].

Telomeres cap each chromosome and protect its integrity. Critically short telomeres lead to genomic instability and cancer development. Telomere length inversely correlates with age, and there is large inter-individual variation of telomere length among people of the same ages [9, 10]. Because telomere length is highly correlated between blood and different tissues in newborns and adults [11-13], telomere length in easily accessible tissues such as blood can serve as a surrogate for telomere length in other tissues. Leukocyte telomere length (LTL) is therefore often used in human population studies to investigate the relationship between a person's telomere length and disease risks [14-18]. A recent study showed that short LTL was associated with poorer RCC-specific survival in two independent patient cohorts [19]. However, there has not been any report of LTL with recurrence in RCC patients.

Recently, there has been increasing use of genetic variants as instrumental variables to estimate LTL and determine its association with disease risks, an approach called Mendelian randomization (MR) [20-23]. There are three assumptions in MR studies: 1) the selected genetic variants are associated with the studied risk factor/biomarker (e.g., LTL); 2) the genetic variants are independent of confounding factors; and 3) the genetic variants only influence disease risk through their effects on the risk factor/biomarker. Large genome-wide association studies (GWAS) have identified at least ten independent genomic regions associated with LTL [24-26]. Single nucleotide polymorphisms (SNPs) in these regions are believed to meet the assumptions of MR studies and have been used as genetic instruments to assess the causal relationship between LTL and diseases risks [20-22, 27-34]. No study has applied an MR approach to study LTL in

RCC prognosis. In this study, for the first time, we evaluated the roles of genetically predicted LTL in the recurrence and survival of RCC patients.

Materials and methods

Study population and data collection

This study consisted of a total of 1,086 patients of European ancestry with histologically confirmed RCC from the University of Texas MD Anderson Cancer Center. The patients were newly diagnosed patients who were treated at MD Anderson. Questionnaire data were obtained via personal interview by trained MD Anderson study interviewers. The questionnaire collects data on demographics, tobacco exposure, occupational history, family history of cancer, medical history, and medication. Clinical and follow-up data were abstracted from medical records. Information include comorbid conditions, pre-treatment performance status, pre-treatment weight loss, location of the primary tumor, tumor size, clinical and pathological stage, histology, tumor grade, treatment type (surgery, cytokine therapy, targeted therapy, chemotherapy, radiotherapy, and other therapy), local recurrence and distant metastasis (date of first recurrence/metastasis), current vital status (date of death and cause of death). The primary clinical endpoints were recurrence (including local recurrence and distant metastasis) and disease-specific survival. Time to recurrence/death was computed from date of surgery to date of last follow-up or recurrence/death.

Genotyping and imputation

Genotyping was performed in the Genotyping Core of MD Anderson Cancer Center using the Illumina HumanHap660W Beadchips and quality control for genotyping has been described previously [35]. Briefly, cases were excluded from analysis if they had genotyping call rates less than 95%, were found to be duplicated samples or known relatives to another sample, were found not to be of European ancestry, and were found to have reported a gender that did not match with X chromosome heterozygosity. We randomly selected 2% of the samples for duplicate genotyping. The concordance of SNP genotype calls was 99.2% for duplicated samples. Individual SNPs with minor allele frequen-

Leukocyte telomere length and prognosis of renal cancer

Table 1. Selected characteristics of the study patients

| Characteristics | N (%) |
|-------------------------------------|-------------|
| Age, Mean (SD) | 59.2 (10.6) |
| Sex | |
| Men | 729 (67.1) |
| Women | 357 (32.9) |
| Smoking status at diagnosis | |
| Never-smoker | 491 (46.1) |
| Former smoker | 428 (40.1) |
| Current smoker | 147 (13.8) |
| BMI at diagnosis, kg/m ² | |
| <25 | 159 (19.9) |
| 25-29.99 (overweight) | 299 (37.5) |
| ≥30 (obese) | 339 (42.5) |
| Histology | |
| Clear Cell | 839 (77.3) |
| Other | 247 (22.7) |
| Clinical Stage | |
| I | 399 (37.5) |
| II | 81 (7.6) |
| III | 292 (27.4) |
| IV | 293 (27.5) |
| Fuhrman Grade | |
| 2 | 358 (36.1) |
| 3 | 421 (42.4) |
| 4 | 214 (21.5) |
| Surgery | |
| Yes | 992 (93.8) |
| No | 66 (6.2) |
| Recurrence | |
| Yes | 440 (40.5) |
| No | 646 (59.5) |
| Survival status | |
| Dead | 467 (43.0) |
| Alive | 619 (57.0) |

cy <1% and call rate <90% were excluded. Imputation was performed using the Michigan Imputation Server (<https://imputationserver.sph.umich.edu/>), an online server that generates phased and imputed genotypes using the Haplotype Reference Consortium (HRC Version r1.1) reference panels. The individual level data of the 10 LTL-associated SNPs were extracted from the genotyped and imputed dataset. Among these SNPs, four SNPs (rs10936599, rs2736100, rs9420907, and rs755017) were directly genotyped, and the other six were imputed with a high imputation accuracy (mean R² of 0.96).

Genetic risk scores (GRS) for LTL

A two-sample MR design was used to assess the associations between genetically predicted LTL and the risk of recurrence and death as described previously [29]. The SNP-LTL associations (estimate for each SNP in **Table 2**) were derived from published genome-wide association studies [24-26] and the SNP-RCC prognosis associations were estimated using individual genotype data in our patient cohort. Genetic risk scores (GRS) calculation for 10 telomere-length associated variants was used according to the following formula.

$$GRS_i = \sum_{j=1}^{10} w_j x_{ij}$$

Where GRS_i is the risk score for individual *i*. *w_j* is the weight or effect coefficient (estimate) for each SNP and *x_{ij}* (*x_{ij}*=0, 1 or 2) is the number of LTL increasing alleles for the *j*-th SNP. Weighted GRS assigned more weight to SNPs with stronger effects and many recent publications have utilized weighted GRS as an instrumental variable to estimate LTL and evaluated its associations with cancer risks and outcomes [27-34].

Statistical analysis

For each LTL-associated SNP, we evaluated its association with the risk of developing recurrence or death by calculating the hazard ratio (HR) and corresponding 95% confidence interval (95% CI) using multivariate Cox proportional hazards model, adjusting for age, smoking status, BMI, stage, grade, and treatment. To analyze the association between GRS and the risk of developing recurrence or death, we dichotomized GRS at the median value or categorized into four groups based on the quartile distribution, and used multivariate Cox proportional hazards model to calculate HR and corresponding 95% CI adjusting for age, smoking status, BMI, stage, grade, and treatment.

Results

Patient characteristics

Table 1 shows the distribution of selected characteristics of the 1,086 RCC patients. The mean age (standard deviation) at diagnosis was 59.2 (10.6) years. There were 729 men (67.1%) and 357 women (32.9%). The majority were never-smokers (45.2%) and former smokers (39.4%), with only 147 (13.5%) current smokers. About 80% of patients were obese

Leukocyte telomere length and prognosis of renal cancer

Table 2. The associations of individual LTL-associated SNPs with the risks of recurrence and death

| SNP ID | Chr. | Position | Gene | Allele* | SNP-LTL | | SNP-Recurrence | | | SNP-Death | | |
|------------|------|-----------|--------|---------|---------|-----------|----------------|------------------|---------|--------------|------------------|---------|
| | | | | | EAF* | β^* | β^{**} | HR** (95% CI) | P value | β^{**} | HR** (95% CI) | P value |
| rs11125529 | 2 | 54475866 | ACYP2 | A/C | 0.14 | 0.07 | -0.15 | 0.86 (0.69-1.08) | 0.206 | -0.14 | 0.87 (0.70-1.09) | 0.222 |
| rs6772228 | 3 | 58376019 | PXK | T/A | 0.94 | 0.04 | -0.32 | 0.72 (0.52-1.02) | 0.062 | -0.02 | 0.98 (0.67-1.42) | 0.901 |
| rs10936599 | 3 | 169492101 | TERC | C/T | 0.75 | 0.10 | -0.05 | 0.95 (0.81-1.12) | 0.542 | -0.16 | 0.85 (0.73-1.00) | 0.047 |
| rs7675998 | 4 | 164007820 | NAF1 | G/A | 0.77 | 0.05 | -0.16 | 0.85 (0.73-0.99) | 0.043 | -0.05 | 0.95 (0.81-1.12) | 0.567 |
| rs2736100 | 5 | 1286516 | TERT | C/A | 0.51 | 0.09 | 0.02 | 1.02 (0.89-1.17) | 0.781 | -0.02 | 0.98 (0.86-1.11) | 0.724 |
| rs9420907 | 10 | 105676465 | OBFC1 | C/A | 0.13 | 0.14 | -0.05 | 0.95 (0.78-1.15) | 0.592 | -0.16 | 0.85 (0.70-1.03) | 0.101 |
| rs3027234 | 17 | 8136092 | CTC1 | C/T | 0.78 | 0.10 | -0.09 | 0.91 (0.76-1.09) | 0.316 | -0.01 | 0.99 (0.84-1.18) | 0.937 |
| rs8105767 | 19 | 22215441 | ZNF208 | G/A | 0.29 | 0.06 | 0.01 | 1.01 (0.88-1.17) | 0.873 | 0.00 | 1.00 (0.87-1.15) | 0.983 |
| rs6028466 | 20 | 38129002 | DHX35 | A/G | 0.06 | 0.06 | -0.03 | 0.97 (0.73-1.29) | 0.834 | -0.02 | 0.98 (0.75-1.29) | 0.898 |
| rs755017 | 20 | 62421622 | ZBTB46 | G/A | 0.12 | 0.02 | 0.08 | 1.08 (0.89-1.31) | 0.436 | -0.10 | 0.91 (0.74-1.10) | 0.321 |

Alleles are short allele/long allele. Short alleles are used as the reference allele and long allele as effect allele. EAF: effect allele frequency; β^ estimates of SNP-LTL association were from published GWAS; β^{**} estimates for SNP-recurrence and SNP-Survival were from this study. **Adjusted by age, smoking status, BMI, histology, stage, grade, and treatment.

Leukocyte telomere length and prognosis of renal cancer

Table 3. GRS predictive of LTL is associated with recurrence in RCC patients

| LTL | No Recurrence N (%) | Recurrence N (%) | Adjusted HR (95% CI)* | P value |
|----------------|------------------------|---------------------|--------------------------|------------|
| Dichotomize | | | | |
| Long | 296 (61.28) | 187 (38.72) | Reference | N/A |
| Short | 275 (56.47) | 212 (43.53) | 1.26 (1.03-1.54) | 0.025 |
| Quartile | | | | |
| 4th (longest) | 151 (63.18) | 88 (36.82) | Reference | N/A |
| 3rd | 145 (59.43) | 99 (40.57) | 1.00 (0.74-1.35) | 0.991 |
| 2nd | 149 (61.32) | 94 (38.68) | 1.01 (0.75-1.36) | 0.967 |
| 1st (shortest) | 126 (51.64) | 118 (48.36) | 1.58 (1.19-2.11) | 0.002 |
| P for trend | | | | 0.002 |

*Adjusted by age, smoking status, BMI, histology, stage, grade, and treatment.

Table 4. GRS predictive of LTL is associated with survival in RCC patients

| LTL | Alive N (%) | Dead N (%) | Adjusted HR (95% CI)* | P value |
|----------------|----------------|---------------|--------------------------|------------|
| Dichotomize | | | | |
| Long | 291 (60.25) | 192 (39.75) | Reference | N/A |
| Short | 270 (55.44) | 217 (44.56) | 1.23 (1.00-1.50) | 0.045 |
| Quartile | | | | |
| 4th (longest) | 151 (63.18) | 88 (36.82) | Reference | N/A |
| 3rd | 140 (57.38) | 104 (42.62) | 1.18 (0.87-1.60) | 0.280 |
| 2nd | 139 (57.20) | 104 (42.80) | 1.24 (0.92-1.68) | 0.157 |
| 1st (shortest) | 131 (53.69) | 113 (46.31) | 1.45 (1.08-1.94) | 0.012 |
| P for trend | | | | 0.013 |

*Adjusted by age, smoking status, BMI, histology, stage, grade, and treatment.

(42.5%) and overweight (37.5%). The distribution of clinical stages was: 399 (37.5%) stage I, 81 (7.6%) stage II, 292 (27.4%) stage III, and 293 stage IV (27.5%). Over three quarters (77.3%) of patients had clear cell RCC. The vast majority (93.8%) of patients received surgical resection of tumors. A total of 440 (40.5%) patients developed recurrence and 467 (43.0%) patients died.

Associations of LTL-associated SNPs and GRS with prognosis of RCC

Table 2 shows the individual associations of 10 LTL-associated SNPs with recurrence and death. Patients carrying the effect allele (longer LTL) of rs7675998 in *NAF1* gene at chromosome 4 exhibited a significantly reduced risk of recurrence (HR=0.85, 95% CI, 0.73-0.99, P=0.043) and the effect allele (long LTL) allele

of rs10936599 in *TERC* gene at chromosome 3 was associated with a significantly reduced risk of death (HR=0.85, 95% CI, 0.73-1.00, P=0.047).

We then constructed a weighted GRS to predict LTL for each patient using the formula described in the Methods. In multivariate Cox analysis adjusting for age, smoking status, BMI, histology, stage, grade, and treatment, lower GRS (shorter LTL) was associated with a significantly increased risk of recurrence (HR=1.32 per SD decrease, 95% CI, 1.02-1.70, P=0.032). When patients were dichotomized into low and high GRS groups based on the median (50th percentile) value of GRS, patients with low GRS (short LTL) had a 1.26-fold (95% CI, 1.03-1.54, P=0.025) increased risk of recurrence compared to those with high GRS (long LTL). When patients were categorized into four groups based on the quartile distribution of GRS, patients with the lowest quartile of GRS (shortest quartile of LTL) were 58% more likely to develop recurrence than those with the highest quartile of GRS (longest quartile of LTL) (HR=1.58, 95% CI, 1.19-2.11 P=0.0016) (**Table 3**).

Likewise, patients with lower GRS (shorter LTL) had a significantly increased risk of death (HR=1.46 per SD decrease, 95% CI, 1.13-1.88, P=0.0038). When patients were dichotomized into low and high GRS groups based on the median value of GRS, patients with low GRS exhibited a 1.23-fold (95% CI, 1.00-1.50, P=0.045) increased risk of death compared to those with high GRS. When patients were categorized into four groups based on the quartile distribution of GRS, the HRs for patients with the lowest quartile of GRS, 2nd quartile, and 3rd quartile of GRS were 1.45 (95% CI, 1.08-1.94, P=0.012), 1.24 (95% CI, 0.92-1.67, P=0.157), and 1.18 (95% CI, 0.87-1.60, P=0.280), respectively (P for trend =0.013), compared to patients with the highest quartile of GRS (**Table 4**).

Discussion

In this study, we applied an MR approach to show that genetically predicted short LTL was an independent predictor of worse prognosis in RCC patients. To our knowledge, this is the first study to evaluate the associations of LTL with the recurrence of RCC and the first one to use MR approach for studying LTL and RCC survival.

Telomeres play a key role in maintaining genomic integrity via protecting chromosomes from degradation, end-to-end fusion, and abnormal recombination [36]. Numerous observational studies have assessed the associations of LTL with the risk of cancers, including RCC [16, 18, 37-41]. Earlier small hospital-based retrospective case control studies reported that short LTL associated with higher risks of RCC [38, 39], which was likely spurious association due to reverse causation, an inherited limitation when evaluating an intermediate biomarker like LTL and cancer risks [37, 42]. Two prospective cohort studies did not find significant associations between pre-diagnostic LTL and RCC risks [16, 41]. In terms of LTL and RCC outcome, an early small study of 105 RCC patients with 28 cancer-specific death found long LTL was associated with an increased risk of cancer-specific death [43]. A recent large study investigated LTL and RCC-specific survival among 684 cases from a population-based U.S. kidney cancer study (USKC) and 241 cases from the prostate, lung, colorectal, and ovarian cancer screening trial (PLCO) and found short LTL was associated with poorer disease-specific survival in both USKC (lowest vs highest quartile: HR: 2.3, 95% CI: 1.2-4.4) and PLCO (HR: 2.4, 95% CI: 1.0-5.4) [19]. No study has specifically evaluated the role of LTL in RCC recurrence. Short LTL has been associated with poor survival in several other cancers [29, 30, 34, 44-47], but a few studies also reported that long LTL was associated with poor prognosis [48-50]. The heterogeneous results between LTL and cancer prognosis may be due to different cancer types, heterogeneous patient population, small sample sizes of most studies, and technical variability. The reproducibility of the popular real-time quantitative PCR technique to measure LTL in human population studies is profoundly impacted by many pre-analytic and analytical factors (e.g., DNA extraction method, storage, and assay conditions) [51-53].

MR study uses genetic variants as a proxy for a risk factor/intermediate biomarker [20-23]. MR study is not susceptible to unmeasured confounding factors and reverse causation typical of retrospective observational studies. In addition, genotyping technology is robust without technical variations. MR study has been increasingly applied in population studies for assessing exposures/biomarkers and disease risks and outcomes and LTL is one of the most commonly studied biomarkers using MR approach [20-22, 27-34]. Two recent large MR studies reported significant associations between longer LTL and increased risks of RCC [21, 27]. Our study is the first to use an MR approach to assess the associations of genetically predicted LTL with RCC recurrence or survival. Consistent with the results of aforementioned observational studies of USKC and PLCO patient populations [19], we found genetically predicted short LTL was associated with a significantly increased risk of death in RCC patients, and for the first time, we also found a significant association between short LTL and recurrence. Our study provided strong evidence for a causal relationship between short LTL and poor RCC prognosis.

The biological mechanisms underlying the associations between short LTL and poor RCC prognosis may lie in two aspects. First, LTL serves as a surrogate for tissue telomere length and short LTL indicates short telomere length in kidney tissues. Numerous studies have shown that somatic telomere shortening increases genomic instability and promote carcinogenesis in mice models [54-58]. In human studies, a high correlation was observed between telomere length in blood and other tissues among newborns and adults [11, 13]. Telomere shortening and telomerase activation were associated with RCC progression and aggressive RCC [59-62]. Therefore, short LTL is a surrogate for short telomeres in kidney tissues, which is a poor prognostic factor. Second, LTL reflects the homeostasis of immune cells and short LTL may therefore indicate increased senescence of immune cells and weakened immunity. In this regard, the peripheral blood leukocytes of colorectal patients with short LTL had higher proportion of CD4(+) T cell and lower proportion of B cell, as well as lower concentration of plasma transforming growth factor- β 1, indicating weakened immune response [44]. Similar observations were made in gastric can-

Leukocyte telomere length and prognosis of renal cancer

cer patients and those with short LTL had an enhanced immunosuppressive status [45]. Therefore, both local and systemic mechanisms may contribute to the observed associations between short LTL and poor prognosis of RCC.

There are both strengths and limitations for our study. This is the first study to report a significant association between short LTL and an increased risk of recurrence in RCC patients. This is also the first MR study of LTL in RCC prognosis. The findings were consistent with a prior observational study [19]. The sample size was large and all the patients were treated at a single institution with consistent treatment and long-term follow-up. In terms of limitations, first, like other MR studies using SNPs as instrument variables, the SNPs used in this study only explain approximately 2% of the variability of LTL. Additional SNPs are desired to enhance instrument strength. Nevertheless, many recent high-impact studies have clearly demonstrated the power of using these SNPs as proxy to assess genetically predicted LTL and various diseases [20-22, 27-34, 63-66]. Second, we only included patients of European ancestry in our analysis due to the small number of minority patients. Future studies are needed to assess the associations of LTL with RCC prognosis in other racial/ethnic groups.

In conclusion, using a Mendelian randomization approach, we found that short LTL is associated with increased risks of recurrence and death for RCC patients of European ancestry. Short LTL may become a biomarker of poor prognosis and facilitate the risk stratification of RCC patients for better-informed clinical management.

Acknowledgements

This study was financially supported by an MD Anderson Cancer Center start-up fund. The funder did not play a role in the design and execution of this study and the writing of the manuscript.

Disclosure of conflict of interest

None.

Address correspondence to: Jian Gu, Department of Epidemiology, The University of Texas MD Anderson

Cancer Center, Houston, TX 77030, USA. E-mail: jiangu@mdanderson.org

References

- [1] Howlader N, Noone AM, Krapcho M, Miller D, Brest A, Yu M, Ruhl J, Tatalovich Z, Mariotto A, Lewis DR, Chen HS, Feuer EJ and Cronin KA (eds). SEER Cancer Statistics Review, 1975-2017, National Cancer Institute. Bethesda, MD, https://seer.cancer.gov/csr/1975_2017/, based on November 2019 SEER data submission, posted to the SEER web site, April 2020, accessed on April 28, 2020.
- [2] Chow WH, Dong LM and Devesa SS. Epidemiology and risk factors for kidney cancer. *Nat Rev Urol* 2010; 7: 245-57.
- [3] Hsieh JJ, Purdue MP, Signoretti S, Swanton C, Albiges L, Schmidinger M, Heng DY, Larkin J and Ficarra V. Renal cell carcinoma. *Nat Rev Dis Primers* 2017; 3: 17009.
- [4] Ravaud A, Motzer RJ, Pandha HS, George DJ, Pantuck AJ, Patel A, Chang YH, Escudier B, Donskov F, Magheli A, Carteni G, Laguerre B, Tomczak P, Breza J, Gerletti P, Lechuga M, Lin X, Martini JF, Ramaswamy K, Casey M, Staehler M and Patard JJ; S-TRAC Investigators. Adjuvant sunitinib in high-risk renal-cell carcinoma after nephrectomy. *N Engl J Med* 2016; 375: 2246-2254.
- [5] Meissner MA, McCormick BZ, Karam JA and Wood CG. Adjuvant therapy for advanced renal cell carcinoma. *Expert Rev Anticancer Ther* 2018; 18: 663-671.
- [6] Bai YY, Li SC, Jia Z, Ding YK, Gu CH and Yang JJ. Adjuvant therapy for locally advanced renal cell carcinoma: a meta-analysis and systematic review. *Urol Oncol* 2018; 36: 79.e1-79.e10.
- [7] Brooks SA, Brannon AR, Parker JS, Fisher JC, Sen O, Kattan MW, Hakimi AA, Hsieh JJ, Choueiri TK, Tamboli P, Maranchie JK, Hinds P, Miller CR, Nielsen ME and Rathmell WK. ClearCode34: a prognostic risk predictor for localized clear cell renal cell carcinoma. *Eur Urol* 2014; 66: 77-84.
- [8] Rini B, Goddard A, Knezevic D, Maddala T, Zhou M, Aydin H, Campbell S, Elson P, Koscielny S, Lopatin M, Svedman C, Martini JF, Williams JA, Verkarre V, Radulescu C, Neuzillet Y, Hemmerlé I, Timsit MO, Tsiatis AC, Bonham M, Lebret T, Mejean A and Escudier B. A 16-gene assay to predict recurrence after surgery in localised renal cell carcinoma: development and validation studies. *Lancet Oncol* 2015; 16: 676-85.
- [9] Aubert G and Lansdorp PM. Telomeres and aging. *Physiol Rev* 2008; 88: 557-79.
- [10] Lansdorp PM, Verwoerd NP, van de Rijke FM, Dragowska V, Little MT, Dirks RW, Raap AK and

Leukocyte telomere length and prognosis of renal cancer

- Tanke HJ. Heterogeneity in telomere length of human chromosomes. *Hum Mol Genet* 1996; 5: 685-91.
- [11] Okuda K, Bardeguet A, Gardner JP, Rodriguez P, Ganesh V, Kimura M, Skurnick J, Awad G and Aviv A. Telomere length in the newborn. *Pediatr Res* 2002; 52: 377-81.
- [12] Friedrich U, Griese E, Schwab M, Fritz P, Thon K and Klotz U. Telomere length in different tissues of elderly patients. *Mech Ageing Dev* 2000; 119: 89-99.
- [13] Daniali L, Benetos A, Susser E, Kark JD, Labat C, Kimura M, Desai K, Granick M and Aviv A. Telomeres shorten at equivalent rates in somatic tissues of adults. *Nat Commun* 2013; 4: 1597.
- [14] Fasching CL. Telomere length measurement as a clinical biomarker of aging and disease. *Crit Rev Clin Lab Sci* 2018; 55: 443-465.
- [15] Mons U, Müezziner A, Schöttker B, Dieffenbach AK, Butterbach K, Schick M, Peasey A, De Vivo I, Trichopoulou A, Boffetta P and Brenner H. Leukocyte telomere length and all-cause, cardiovascular disease, and cancer mortality: results from individual-participant-data meta-analysis of 2 large prospective cohort studies. *Am J Epidemiol* 2017; 185: 1317-1326.
- [16] Weischer M, Nordestgaard BG, Cawthon RM, Freiberg JJ, Tybjaerg-Hansen A and Bojesen SE. Short telomere length, cancer survival, and cancer risk in 47102 individuals. *J Natl Cancer Inst* 2013; 105: 459-68.
- [17] Rode L, Nordestgaard BG and Bojesen SE. Long telomeres and cancer risk among 95 568 individuals from the general population. *Int J Epidemiol* 2016; 45: 1634-1643.
- [18] Zhang X, Zhao Q, Zhu W, Liu T, Xie SH, Zhong LX, Cai YY, Li XN, Liang M, Chen W, Hu QS and Zhang B. The association of telomere length in peripheral blood cells with cancer risk: a systematic review and meta-analysis of prospective studies. *Cancer Epidemiol Biomarkers Prev* 2017; 26: 1381-1390.
- [19] Callahan CL, Schwartz K, Ruterbusch JJ, Shuch B, Graubard BI, Lan Q, Cawthon R, Baccarelli AA, Chow WH, Rothman N, Hofmann JN and Purdue MP. Leukocyte telomere length and renal cell carcinoma survival in two studies. *Br J Cancer* 2017; 117: 752-755.
- [20] Zhang C, Doherty JA, Burgess S, Hung RJ, Lindström S, Kraft P, Gong J, Amos CI, Sellers TA, Monteiro AN, Chenevix-Trench G, Bickeböller H, Risch A, Brennan P, McKay JD, Houlston RS, Landi MT, Timofeeva MN, Wang Y, Heinrich J, Kote-Jarai Z, Eeles RA, Muir K, Wiklund F, Grönberg H, Berndt SI, Chanock SJ, Schumacher F, Haiman CA, Henderson BE, Amin Al Olama A, Andrulis IL, Hopper JL, Chang-Claude J, John EM, Malone KE, Gammon MD, Ursin G, Whittemore AS, Hunter DJ, Gruber SB, Knight JA, Hou L, Le Marchand L, Newcomb PA, Hudson TJ, Chan AT, Li L, Woods MO, Ahsan H and Pierce BL; GECCO and GAME-ON Network: CORECT, DRIVE, ELLIPSE, FOCI, and TRICL. Genetic determinants of telomere length and risk of common cancers: a Mendelian randomization study. *Hum Mol Genet* 2015; 24: 5356-66.
- [21] Telomeres Mendelian Randomization Collaboration, Haycock PC, Burgess S, Nounu A, Zheng J, Okoli GN, Bowden J, Wade KH, Timpson NJ, Evans DM, Willeit P, Aviv A, Gaunt TR, Hemani G, Mangino M, Ellis HP, Kurian KM, Pooley KA, Eeles RA, Lee JE, Fang S, Chen WV, Law MH, Bowdler LM, Iles MM, Yang Q, Worrall BB, Markus HS, Hung RJ, Amos CI, Spurdle AB, Thompson DJ, O'Mara TA, Wolpin B, Amundadottir L, Stolzenberg-Solomon R, Trichopoulou A, Onland-Moret NC, Lund E, Duell EJ, Canzian F, Severi G, Overvad K, Gunter MJ, Tumino R, Svenson U, van Rij A, Baas AF, Bown MJ, Samani NJ, van t'Hof FNG, Tromp G, Jones GT, Kuivaniemi H, Elmore JR, Johansson M, McKay J, Scelo G, Carreras-Torres R, Gaborieau V, Brennan P, Bracci PM, Neale RE, Olson SH, Gallinger S, Li D, Petersen GM, Risch HA, Klein AP, Han J, Abnet CC, Freedman ND, Taylor PR, Maris JM, Aben KK, Kiemeny LA, Vermeulen SH, Wiencke JK, Walsh KM, Wrensch M, Rice T, Turnbull C, Litchfield K, Paternoster L, Standl M, Abecasis GR, SanGiovanni JP, Li Y, Mijatovic V, Sapkota Y, Low SK, Zondervan KT, Montgomery GW, Nyholt DR, van Heel DA, Hunt K, Arking DE, Ashar FN, Sotoodehnia N, Woo D, Rosand J, Comeau ME, Brown WM, Silverman EK, Hokanson JE, Cho MH, Hui J, Ferreira MA, Thompson PJ, Morrison AC, Felix JF, Smith NL, Christiano AM, Petukhova L, Betz RC, Fan X, Zhang X, Zhu C, Langefeld CD, Thompson SD, Wang F, Lin X, Schwartz DA, Fingerlin T, Rotter JI, Cotch MF, Jensen RA, Munz M, Dommisch H, Schaefer AS, Han F, Ollila HM, Hillary RP, Albagha O, Ralston SH, Zeng C, Zheng W, Shu XO, Reis A, Uebe S, Hüffmeier U, Kawamura Y, Otowa T, Sasaki T, Hibberd ML, Davila S, Xie G, Siminovitsh K, Bei JX, Zeng YX, Försti A, Chen B, Landi S, Franke A, Fischer A, Ellinghaus D, Flores C, Noth I, Ma SF, Foo JN, Liu J, Kim JW, Cox DG, Delattre O, Mirabeau O, Skibola CF, Tang CS, Garcia-Barcelo M, Chang KP, Su WH, Chang YS, Martin NG, Gordon S, Wade TD, Lee C, Kubo M, Cha PC, Nakamura Y, Levy D, Kimura M, Hwang SJ, Hunt S, Spector T, Soranzo N, Manichaikul AW, Barr RG, Kahali B, Speliotes E, Yerges-Armstrong LM, Cheng CY, Jonas JB, Wong TY, Fogh I, Lin K, Powell JF, Rice K, Relton CL, Martin RM and Davey Smith G. Association Between telomere length and

Leukocyte telomere length and prognosis of renal cancer

- risk of cancer and non-neoplastic diseases: a mendelian randomization study. *JAMA Oncol* 2017; 3: 636-651.
- [22] Pierce BL, Kraft P and Zhang C. Mendelian randomization studies of cancer risk: a literature review. *Curr Epidemiol Rep* 2018; 5: 184-196.
- [23] Lawlor DA, Harbord RM, Sterne JA, Timpson N and Davey Smith G. Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. *Stat Med* 2008; 27: 1133-1163.
- [24] Pooley KA, Bojesen SE, Weischer M, Nielsen SF, Thompson D, Amin Al Olama A, Michailidou K, Tyrer JP, Benlloch S, Brown J, Audley T, Luben R, Khaw KT, Neal DE, Hamdy FC, Donovan JL, Kote-Jarai Z, Baynes C, Shah M, Bolla MK, Wang Q, Dennis J, Dicks E, Yang R, Rudolph A, Schildkraut J, Chang-Claude J, Burwinkel B, Chenevix-Trench G, Pharoah PD, Berchuck A, Eeles RA, Easton DF, Dunning AM and Nordestgaard BG. A genome-wide association scan (GWAS) for mean telomere length within the COGS project: identified loci show little association with hormone-related cancer risk. *Hum Mol Genet* 2013; 22: 5056-64.
- [25] Mangino M, Hwang SJ, Spector TD, Hunt SC, Kimura M, Fitzpatrick AL, Christiansen L, Petersen I, Elbers CC, Harris T, Chen W, Srinivasan SR, Kark JD, Benetos A, El Shamieh S, Visvikis-Siest S, Christensen K, Berenson GS, Valdes AM, Viñuela A, Garcia M, Arnett DK, Broeckel U, Province MA, Pankow JS, Kammerer C, Liu Y, Nalls M, Tishkoff S, Thomas F, Ziv E, Psaty BM, Bis JC, Rotter JI, Taylor KD, Smith E, Schork NJ, Levy D and Aviv A. Genome-wide meta-analysis points to CTC1 and ZNF676 as genes regulating telomere homeostasis in humans. *Hum Mol Genet* 2012; 21: 5385-94.
- [26] Codd V, Nelson CP, Albrecht E, Mangino M, Deelen J, Buxton JL, Hottenga JJ, Fischer K, Esko T, Surakka I, Broer L, Nyholt DR, Mateo Leach I, Salo P, Hägg S, Matthews MK, Palmen J, Norata GD, O'Reilly PF, Saleheen D, Amin N, Balmforth AJ, Beekman M, de Boer RA, Böhringer S, Braund PS, Burton PR, de Craen AJ, Denniff M, Dong Y, Douroudis K, Dubinina E, Eriksson JG, Garlaschelli K, Guo D, Hartikainen AL, Henders AK, Houwing-Duistermaat JJ, Kananen L, Karssen LC, Kettunen J, Klopp N, Lagou V, van Leeuwen EM, Madden PA, Mägi R, Magnusson PK, Männistö S, McCarthy MI, Medland SE, Mihailov E, Montgomery GW, Oostra BA, Palotie A, Peters A, Pollard H, Pouta A, Prokopenko I, Ripatti S, Salomaa V, Surchiman HE, Valdes AM, Verweij N, Viñuela A, Wang X, Wichmann HE, Widen E, Willemsen G, Wright MJ, Xia K, Xiao X, van Veldhuisen DJ, Catapano AL, Tobin MD, Hall AS, Blakemore AI, van Gilst WH, Zhu H; CARDIoGRAM consortium, Erdmann J, Reilly MP, Kathiresan S, Schunkert H, Talmud PJ, Pedersen NL, Perola M, Ouwehand W, Kaprio J, Martin NG, van Duijn CM, Hovatta I, Gieger C, Metspalu A, Boomsma DI, Jarvelin MR, Slagboom PE, Thompson JR, Spector TD, van der Harst P and Samani NJ. Identification of seven loci affecting mean telomere length and their association with disease. *Nat Genet* 2013; 45: 422-7, 427, e1-2.
- [27] Machiela MJ, Hofmann JN, Carreras-Torres R, Brown KM, Johansson M, Wang Z, Foll M, Li P, Rothman N, Savage SA, Gaborieau V, McKay JD, Ye Y, Henrion M, Bruinsma F, Jordan S, Severi G, Hveem K, Vatten LJ, Fletcher T, Koppova K, Larsson SC, Wolk A, Banks RE, Selby PJ, Easton DF, Pharoah P, Andreotti G, Freeman LEB, Koutros S, Albanes D, Mannisto S, Weinstein S, Clark PE, Edwards TE, Lipworth L, Gapstur SM, Stevens VL, Carol H, Freedman ML, Pomerantz MM, Cho E, Kraft P, Preston MA, Wilson KM, Gaziano JM, Sesso HS, Black A, Freedman ND, Huang WY, Anema JG, Kahnoski RJ, Lane BR, Noyes SL, Petillo D, Colli LM, Sampson JN, Besse C, Blanche H, Boland A, Burdette L, Prokhorchouk E, Skryabin KG, Yeager M, Mijuskovic M, Ognjanovic M, Foretova L, Holcatova I, Janout V, Mates D, Mukeriya A, Rascu S, Zaridze D, Bencko V, Cybulski C, Fabianova E, Jinga V, Lissowska J, Lubinski J, Navratilova M, Rudnai P, Szeszenia-Dabrowska N, Benhamou S, Cancel-Tassin G, Cussenot O, Bueno-de-Mesquita HB, Canzian F, Duell EJ, Ljungberg B, Sitaram RT, Peters U, White E, Anderson GL, Johnson L, Luo J, Buring J, Lee IM, Chow WH, Moore LE, Wood C, Eisen T, Larkin J, Choueiri TK, Lathrop GM, Teh BT, Deleuze JF, Wu X, Houlston RS, Brennan P, Chanock SJ, Scelo G and Purdue MP. Genetic variants related to longer telomere length are associated with increased risk of renal cell carcinoma. *Eur Urol* 2017; 72: 747-754.
- [28] Xu Y, Xu J, Chancoco H, Huang M, Torres KE and Gu J. Long leukocyte telomere length is associated with increased risks of soft tissue sarcoma: a mendelian randomization study. *Cancers (Basel)* 2020; 12: 594.
- [29] Xu J, Chang WS, Tsai CW, Bau DT, Xu Y, Davis JW, Thompson TC, Logothetis CJ and Gu J. Leukocyte telomere length is associated with aggressive prostate cancer in localized prostate cancer patients. *EBioMedicine* 2020; 52: 102616.
- [30] Rachakonda S, Srinivas N, Mahmoudpour SH, Garcia-Casado Z, Requena C, Traves V, Soriano V, Cardelli M, Pjanova D, Molven A, Gruis N, Nagore E and Kumar R. Telomere length and survival in primary cutaneous melanoma patients. *Sci Rep* 2018; 8: 10947.

Leukocyte telomere length and prognosis of renal cancer

- [31] Campa D, Matarazzi M, Greenhalf W, Bijlsma M, Saum KU, Pasquali C, van Laarhoven H, Szentesi A, Federici F, Vodicka P, Funel N, Pezzilli R, Bueno-de-Mesquita HB, Vodickova L, Basso D, Obazee O, Hackert T, Soucek P, Cuk K, Kaiser J, Sperti C, Lovecek M, Capurso G, Mohelnikova-Duchonova B, Khaw KT, König AK, Kupcinskas J, Kaaks R, Bambi F, Archibugi L, Mambri A, Cavestro GM, Landi S, Hegyi P, Izbicki JR, Gioffreda D, Zambon CF, Tavano F, Talar-Wojnarowska R, Jamroziak K, Key TJ, Fave GD, Strobel O, Jonaitis L, Andriulli A, Lawlor RT, Pirozzi F, Katzke V, Valsuani C, Vashist YK, Brenner H and Canzian F. Genetic determinants of telomere length and risk of pancreatic cancer: a PANDORA study. *Int J Cancer* 2019; 144: 1275-1283.
- [32] Cao X, Huang M, Zhu M, Fang R, Ma Z, Jiang T, Dai J, Ma H, Jin G, Shen H, Du J, Xu L and Hu Z. Mendelian randomization study of telomere length and lung cancer risk in East Asian population. *Cancer Med* 2019; 8: 7469-7476.
- [33] Gramatges MM, Morton LM, Yasui Y, Arnold MA, Neglia JP, Leisenring WM, Machiela MJ, Dagnall CL, Chanock SJ, Armstrong GT, Robison LL, Bhatia S and Lupo PJ. Telomere length-associated genetic variants and the risk of thyroid cancer in survivors of childhood cancer: a report from the childhood cancer survivor study (CCSS). *Cancer Epidemiol Biomarkers Prev* 2019; 28: 417-419.
- [34] Song N, Li Z, Qin N, Howell CR, Wilson CL, Easton J, Mulder HL, Edmonson MN, Rusch MC, Zhang J, Hudson MM, Yasui Y, Robison LL, Ness KK and Wang Z. Shortened leukocyte telomere length associates with an increased prevalence of chronic health conditions among survivors of childhood cancer: a report from the St. Jude lifetime cohort. *Clin Cancer Res* 2020; 26: 2362-2371.
- [35] Wu X, Scelo G, Purdue MP, Rothman N, Johansson M, Ye Y, Wang Z, Zelenika D, Moore LE, Wood CG, Prokhortchouk E, Gaborieau V, Jacobs KB, Chow WH, Toro JR, Zaridze D, Lin J, Lubinski J, Trubicka J, Szeszenia-Dabrowska N, Lissowska J, Rudnai P, Fabianova E, Mates D, Jinga V, Bencko V, Slamova A, Holcatova I, Navratilova M, Janout V, Boffetta P, Colt JS, Davis FG, Schwartz KL, Banks RE, Selby PJ, Harnden P, Berg CD, Hsing AW, Grubb RL 3rd, Boeing H, Vineis P, Clavel-Chapelon F, Palli D, Tumino R, Krogh V, Panico S, Duell EJ, Quirós JR, Sanchez MJ, Navarro C, Ardanaz E, Dorronsoro M, Khaw KT, Allen NE, Bueno-de-Mesquita HB, Peeters PH, Trichopoulos D, Linseisen J, Ljungberg B, Overvad K, Tjønneland A, Romieu I, Riboli E, Stevens VL, Thun MJ, Diver WR, Gapstur SM, Pharoah PD, Easton DF, Albanes D, Virtamo J, Vatten L, Hveem K, Fletcher T, Koppova K, Cussenot O, Cancel-Tassin G, Benhamou S, Hildebrandt MA, Pu X, Foglio M, Lechner D, Hutchinson A, Yeager M, Fraumeni JF Jr, Lathrop M, Skryabin KG, McKay JD, Gu J, Brennan P and Chanock SJ. A genome-wide association study identifies a novel susceptibility locus for renal cell carcinoma on 12p11.23. *Hum Mol Genet* 2012; 21: 456-62.
- [36] Blackburn EH. Telomeres and telomerase: their mechanisms of action and the effects of altering their functions. *FEBS Lett* 2005; 579: 859-62.
- [37] Wentzensen IM, Mirabello L, Pfeiffer RM and Savage SA. The association of telomere length and cancer: a meta-analysis. *Cancer Epidemiol Biomarkers Prev* 2011; 20: 1238-50.
- [38] Wu X, Amos CI, Zhu Y, Zhao H, Grossman BH, Shay JW, Luo S, Hong WK and Spitz MR. Telomere dysfunction: a potential cancer predisposition factor. *J Natl Cancer Inst* 2003; 95: 1211-8.
- [39] Shao L, Wood CG, Zhang D, Tannir NM, Matin S, Dinney CP and Wu X. Telomere dysfunction in peripheral lymphocytes as a potential predisposition factor for renal cancer. *J Urol* 2007; 178: 1492-6.
- [40] Hofmann JN, Baccarelli A, Schwartz K, Davis FG, Ruterbusch JJ, Hoxha M, McCarthy BJ, Savage SA, Wacholder S, Rothman N, Graubard BI, Colt JS, Chow WH and Purdue MP. Risk of renal cell carcinoma in relation to blood telomere length in a population-based case-control study. *Br J Cancer* 2011; 105: 1772-5.
- [41] Hofmann JN, Lan Q, Cawthon R, Hosgood HD 3rd, Shuch B, Moore LE, Rothman N, Chow WH and Purdue MP. A prospective study of leukocyte telomere length and risk of renal cell carcinoma. *Cancer Epidemiol Biomarkers Prev* 2013; 22: 997-1000.
- [42] Pooley KA, Sandhu MS, Tyrer J, Shah M, Driver KE, Luben RN, Bingham SA, Ponder BA, Pharoah PD, Khaw KT, Easton DF and Dunning AM. Telomere length in prospective and retrospective cancer case-control studies. *Cancer Res* 2010; 70: 3170-6.
- [43] Svenson U, Ljungberg B and Roos G. Telomere length in peripheral blood predicts survival in clear cell renal cell carcinoma. *Cancer Res* 2009; 69: 2896-901.
- [44] Chen Y, Qu F, He X, Bao G, Liu X, Wan S and Xing J. Short leukocyte telomere length predicts poor prognosis and indicates altered immune functions in colorectal cancer patients. *Ann Oncol* 2014; 25: 869-76.
- [45] Qu F, Li R, He X, Li Q, Xie S, Gong L, Ji G, Lu J and Bao G. Short telomere length in peripheral blood leukocyte predicts poor prognosis and indicates an immunosuppressive phenotype in gastric cancer patients. *Mol Oncol* 2015; 9: 727-39.

Leukocyte telomere length and prognosis of renal cancer

- [46] Kachuri L, Helby J, Bojesen SE, Christiani DC, Su L, Wu X, Tardón A, Fernández-Tardón G, Field JK, Davies MP, Chen C, Goodman GE, Shepherd FA, Leighl NB, Tsao MS, Brhane Y, Brown MC, Boyd K, Shepshelovich D, Sun L, Amos CI, Liu G and Hung RJ. Investigation of leukocyte telomere length and genetic variants in chromosome 5p15.33 as prognostic markers in lung cancer. *Cancer Epidemiol Biomarkers Prev* 2019; 28: 1228-1237.
- [47] Hamada T, Yuan C, Bao Y, Zhang M, Khalaf N, Babic A, Morales-Oyarvide V, Cochrane BB, Gaziano JM, Giovannucci EL, Kraft P, Manson JE, Ng K, Nowak JA, Rohan TE, Sesso HD, Stampfer MJ, Amundadottir LT, Fuchs CS, De Vivo I, Ogino S and Wolpin BM. Prediagnostic leukocyte telomere length and pancreatic cancer survival. *Cancer Epidemiol Biomarkers Prev* 2019; 28: 1868-1875.
- [48] Kim ES, Ye Y, Vaporciyan AA, Xing J, Huang M, Gu J, Roth JA, Lippman SM and Wu X. Telomere length and recurrence risk after curative resection in patients with early-stage non-small-cell lung cancer: a prospective cohort study. *J Thorac Oncol* 2015; 10: 302-8.
- [49] Svenson U, Öberg Å, Stenling R, Palmqvist R and Roos G. Breast cancer survival is associated with telomere length in peripheral blood cells. *Cancer Res* 2008; 68: 3618-23.
- [50] Luo X, Sturgis EM, Yang Z, Sun Y, Wei P, Liu Z, Wei Q and Li G. Lymphocyte telomere length predicts clinical outcomes of HPV-positive oropharyngeal cancer patients after definitive radiotherapy. *Carcinogenesis* 2019; 40: 735-741.
- [51] Aviv A. The epidemiology of human telomeres: faults and promises. *J Gerontol A Biol Sci Med Sci* 2008; 63: 979-83.
- [52] Martin-Ruiz CM, Baird D, Roger L, Boukamp P, Kronic D, Cawthon R, Dokter MM, van der Harst P, Bekaert S, de Meyer T, Roos G, Svenson U, Codd V, Samani NJ, McGlynn L, Shiels PG, Pooley KA, Dunning AM, Cooper R, Wong A, Kingston A and von Zglinicki T. Reproducibility of telomere length assessment: an international collaborative study. *Int J Epidemiol* 2015; 44: 1673-83.
- [53] Dagnall CL, Hicks B, Teshome K, Hutchinson AA, Gadalla SM, Khincha PP, Yeager M and Savage S. Effect of pre-analytic variables on the reproducibility of qPCR relative telomere length measurement. *PLoS One* 2017; 12: e0184098.
- [54] Blasco MA, Lee HW, Hande MP, Samper E, Lansdorp PM, DePinho RA and Greider C. Telomere shortening and tumor formation by mouse cells lacking telomerase RNA. *Cell* 1997; 91: 25-34.
- [55] Chin L, Artandi SE, Shen Q, Tam A, Lee SL, Gottlieb GJ, Greider CW and DePinho RA. p53 deficiency rescues the adverse effects of telomere loss and cooperates with telomere dysfunction to accelerate carcinogenesis. *Cell* 1999; 97: 527-38.
- [56] Hemann MT, Strong MA, Hao LY and Greider CW. The shortest telomere, not average telomere length, is critical for cell viability and chromosome stability. *Cell* 2001; 107: 67-77.
- [57] Nakamura TM, Cooper JP and Cech TR. Two modes of survival of fission yeast without telomerase. *Science* 1998; 282: 493-6.
- [58] Naito T, Matsuura A and Ishikawa F. Circular chromosome formation in a fission yeast mutant defective in two ATM homologues. *Nat Genet* 1998; 20: 203-6.
- [59] Fan Y, Liu Z, Fang X, Ge Z, Ge N, Jia Y, Sun P, Lou F, Björkholm M, Gruber A, Ekman P and Xu D. Differential expression of full-length telomerase reverse transcriptase mRNA and telomerase activity between normal and malignant renal tissues. *Clin Cancer Res* 2005; 11: 4331-7.
- [60] Dahse R, Fiedler W, Junker K, Schlichter A, Schubert J and Claussen U. Telomerase activity and telomere lengths: alterations in renal cell carcinomas. *Kidney Int* 1999; 56: 1289-90.
- [61] Gisselsson D, Gorunova L, Höglund M, Mandahl N and Elfving P. Telomere shortening and mitotic dysfunction generate cytogenetic heterogeneity in a subgroup of renal cell carcinomas. *Br J Cancer* 2004; 91: 327-32.
- [62] Stewénius Y, Jin Y, Øra I, de Kraker J, Bras J, Frigyesi A, Alumets J, Sandstedt B, Meeker AK and Gisselsson D. Defective chromosome segregation and telomere dysfunction in aggressive Wilms' tumors. *Clin Cancer Res* 2007; 13: 6593-602.
- [63] Zhan Y, Karlsson IK, Karlsson R, Tillander A, Reynolds CA, Pedersen NL and Hägg S. Exploring the causal pathway from telomere length to coronary heart disease: a network mendelian randomization study. *Circ Res* 2017; 121: 214-219.
- [64] Kuo CL, Pilling LC, Kuchel GA, Ferrucci L and Melzer D. Telomere length and aging-related outcomes in humans: a Mendelian randomization study in 261,000 older participants. *Aging Cell* 2019; 18: e13017.
- [65] Kachuri L, Saarela O, Bojesen SE, Davey Smith G, Liu G, Landi MT, Caporaso NE, Christiani DC, Johansson M, Panico S, Overvad K, Trichopoulos A, Vineis P, Scelo G, Zaridze D, Wu X, Albanes D, Diergaarde B, Lagiou P, Macfarlane GJ, Aldrich MC, Tardón A, Rennert G, Olshan AF, Weessler MC, Chen C, Goodman GE, Doherty JA, Ness AR, Bickeböller H, Wichmann HE, Risch A, Field JK, Teare MD, Kiemeny LA, van der Heijden EHF, Carroll JC, Haugen A, Zienolddiny S, Skaug V, Wünsch

Leukocyte telomere length and prognosis of renal cancer

Filho V, Tajara EH, Ayoub Moysés R, Daumas Nunes F, Lam S, Eluf-Neto J, Lacko M, Peters WHM, Le Marchand L, Duell EJ, Andrew AS, Franceschi S, Schabath MB, Manjer J, Arnold S, Lazarus P, Mukeriya A, Swiatkowska B, Janout V, Holcatova I, Stojic J, Mates D, Lissowska J, Boccia S, Lesseur C, Zong X, McKay JD, Brennan P, Amos CI and Hung RJ. Mendelian randomization and mediation analysis of leukocyte telomere length and risk of lung and head and neck cancers. *Int J Epidemiol* 2019; 48: 751-766.

[66] Demanelis K, Tong L and Pierce BL. Genetically increased telomere length and Aging-related Traits in the UK biobank. *J Gerontol A Biol Sci Med Sci* 2019: glz240.